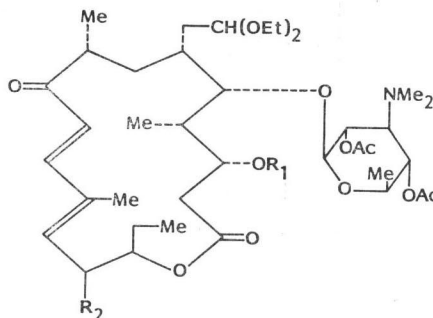


SYNTHESES OF 23-C-SUBSTITUTED  
DERIVATIVES OF  
MYCAMINOSYL TYLONOLIDE

Sir:

In previous papers<sup>1-4</sup> we have reported the syntheses of several types of active derivatives of mycaminosyl tylonolide (MT) and other related compounds which have various substituents at the C-23. This communication describes another kind of derivation at the C-23 of MT, that is, 23-C-alkylation or -arylation. 2',4'-Di-*O*-acetylmycaminosyl tylonolide diethyl acetal (**1**)<sup>5</sup> was treated with *t*-butylchlorodimethylsilane in the presence of imidazole in DMF at 75°C, overnight. The resulting 3,23-bis(*O*-*tert*-butyldimethylsilyl) derivative (**2**) obtained in 75% yield,  $[\alpha]_D^{25} -11^\circ$  (*c* 1, CHCl<sub>3</sub>), was treated with NBu<sub>4</sub>F in oxolane (1 mol equivalent, room temperature, 1 hour) to give the 3-*O*-silyl derivative (**3**) (95%),  $[\alpha]_D^{25} -10^\circ$  (*c* 1, CHCl<sub>3</sub>). Oxidation of **3** with DMSO-benzene (1:1) in the presence of pyridinium trifluoroacetate and dicyclohexylcarbodiimide gave the corresponding 23-aldehyde (**4**) (86%):  $[\alpha]_D^{25} -40^\circ$  (*c* 1, CHCl<sub>3</sub>); <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  9.67 (1H d, 23-H). Treatment of **4** with MeMgBr in oxolane (-78°C, 1.5 hours, then room temperature, 1 hour) gave the (23*S*)-23-*C*-methyl (**5a**) and (23*R*)-23-*C*-methyl (**5b**) derivatives in 36 and 10% yields. **5a**:  $[\alpha]_D^{25} -10^\circ$  (*c* 1, CHCl<sub>3</sub>); <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  1.12 (d, 24-CH<sub>3</sub>). **5b**:  $[\alpha]_D^{25} +11^\circ$  (*c* 1, CHCl<sub>3</sub>); <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  1.22 (d, 24-CH<sub>3</sub>). Compounds **5a** and **5b** were deacetylated (in MeOH, 50°C, overnight), and deblocked with 0.5 M HCl in CH<sub>3</sub>CN (37°C, overnight) to give the final products, **6a** (74%):  $[\alpha]_D^{25} -10^\circ$  (*c* 1, CHCl<sub>3</sub>) and **6b** (78%):  $[\alpha]_D^{25} +16^\circ$  (*c* 1, CHCl<sub>3</sub>). <sup>1</sup>H NMR (CDCl<sub>3</sub>): **6a**:  $\delta$  1.15 (24-CH<sub>3</sub>), **6b**:  $\delta$  1.24 (24-CH<sub>3</sub>). Absolute configurations at C-23 of these products were determined<sup>6</sup> by the nuclear Overhauser effect difference spectroscopy. Irradiation of **6a** at the  $\delta$  1.15 (24-CH<sub>3</sub>) caused pronounced positive signal enhancements of 13-H and 14-H, whereas irradiation of **6b** at the  $\delta$  1.24 (24-CH<sub>3</sub>) caused similar enhancements of 13-H and 15-H. These results led to the conclusion<sup>6</sup>, from the stereochemical requirements, that the absolute configurations at C-23 in **6a** and **6b** should be specified L and D, respectively as shown in Fischer projection (see Fig. 2).

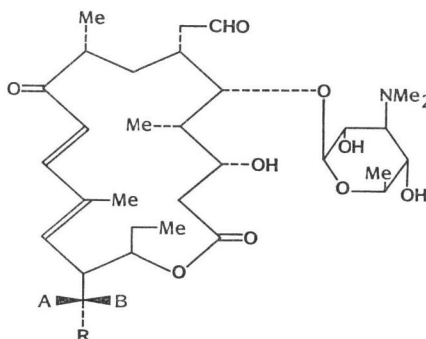
Fig. 1.



<b>1</b>	R <sub>1</sub> =H	R <sub>2</sub> =CH <sub>2</sub> OH
<b>2</b>	R <sub>1</sub> =SiMe <sub>2</sub> ( <i>t</i> Bu)	R <sub>2</sub> =CH <sub>2</sub> OSiMe <sub>2</sub> ( <i>t</i> Bu)
<b>3</b>	R <sub>1</sub> =SiMe <sub>2</sub> ( <i>t</i> Bu)	R <sub>2</sub> =CH <sub>2</sub> OH
<b>4</b>	R <sub>1</sub> =SiMe <sub>2</sub> ( <i>t</i> Bu)	R <sub>2</sub> =CHO
<b>5a</b>	R <sub>1</sub> =SiMe <sub>2</sub> ( <i>t</i> Bu)	R <sub>2</sub> =( <i>S</i> )CH(OH)Me
<b>5b</b>	R <sub>1</sub> =SiMe <sub>2</sub> ( <i>t</i> Bu)	R <sub>2</sub> =( <i>R</i> )CH(OH)Me

<sup>t</sup>Bu: *tert*-Butyl.

Fig. 2.



	a.c.*	A	B	R
<b>6a</b>	<i>S</i>	OH	H	Me
<b>6b</b>	<i>R</i>	H	OH	Me
<b>7a</b>	<i>S</i>	OH	H	Et
<b>7b</b>	<i>R</i>	H	OH	Et
<b>8a</b>	<i>S</i>	OH	H	Bu
<b>8b</b>	<i>R</i>	H	OH	Bu
<b>9a</b>	<i>R</i>	OH	H	C <sub>6</sub> H <sub>5</sub>
<b>9b</b>	<i>S</i>	H	OH	C <sub>6</sub> H <sub>5</sub>
<b>10a</b>	<i>S</i>	OH	H	CH <sub>2</sub> CH=CH <sub>2</sub>
<b>10b</b>	<i>R</i>	H	OH	CH <sub>2</sub> CH=CH <sub>2</sub>
<b>11a</b>	<i>S</i>	OH	H	CH=CH <sub>2</sub>
<b>11b</b>	<i>R</i>	H	OH	CH=CH <sub>2</sub>
<b>12a</b>	<i>R</i>	OH	H	C≡CH
<b>12b</b>	<i>S</i>	H	OH	C≡CH

\* Absolute configuration at C-23 by CAHN-INGOLD-PRELOG specifications.

Table 1. Antibacterial activity (MIC,  $\mu\text{g/ml}$ ) of derivatives of mycaminosyl tylonolide (MT).

Test organisms	MT	6a	6b	7a	7b	8a	8b
<i>Staphylococcus aureus</i> 193	1.56	1.56	0.78	0.39	0.39	<0.2	<0.2
<i>S. aureus</i> EMf	50	>100	>100	>100	100	25	50
<i>S. aureus</i> 209P	1.56	0.78	0.39	0.39	0.39	<0.2	<0.2
<i>S. aureus</i> MS 9610	>100	>100	>100	>100	>100	>100	>100
<i>S. aureus</i> MS 9351	>100	>100	>100	>100	>100	>100	>100
<i>S. aureus</i> MS 9861	1.56	1.56	0.78	0.78	0.78	0.39	0.39
<i>S. aureus</i> MS 10225	3.12	3.12	1.56	1.56	0.78	0.39	0.78
<i>S. aureus</i> MS 10246	>100	>100	>100	>100	>100	>100	>100
<i>S. aureus</i> Smith	1.56	1.56	1.56	0.78	0.78	0.39	0.39
<i>Micrococcus luteus</i> PCI 1001	<0.2	<0.2	<0.2	<0.2	<0.2	<0.2	<0.2
<i>Bacillus subtilis</i> NRRL B-558	3.12	0.78	0.78	0.78	0.39	0.39	<0.2
<i>Corynebacterium bovis</i> 1810	3.12	3.12	0.78	3.12	0.78	<0.2	<0.2
<i>Escherichia coli</i> NIHJ	12.5	6.25	3.12	3.12	3.12	6.25	3.12
<i>E. coli</i> K-12	25	25	12.5	12.5	12.5	25	12.5
<i>E. coli</i> K-12 R-5	50	50	25	25	12.5	25	12.5
<i>E. coli</i> K-12 ML 1629	100	>100	100	100	100	100	50
<i>E. coli</i> K-12 ML 1410	>100	>100	100	100	100	100	50
<i>E. coli</i> K-12 ML 1410 R81	100	>100	100	100	100	100	100
<i>E. coli</i> K-12 LA290 R55	100	50	50	50	50	25	25
<i>Klebsiella pneumoniae</i> PCI 602	3.12	3.12	3.12	12.5	6.25	3.12	6.25
<i>Shigella dysenteriae</i> JS 11910	1.56	0.78	0.78	1.56	1.56	0.78	0.78
<i>Salmonella enteritidis</i> 1891	3.12	3.12	3.12	3.12	3.12	3.12	3.12
<i>S. typhi</i> T-63	100	100	50	100	50	50	50
<i>Enterobacter aerogenes</i> ATCC 13048	100	100	100	100	100	100	50
<i>Providencia</i> sp. Pv 16	>100	>100	>100	>100	>100	>100	100
<i>Serratia marcescens</i>	50	25	12.5	50	50	50	12.5
<i>Proteus vulgaris</i> OX-19	50	50	25	50	50	25	25
<i>Pseudomonas aeruginosa</i> A3	25	12.5	12.5	6.25	6.25	50	6.25
Geometrical mean	17.25	15.62	10.78	12.50	10.26	8.03	6.12

LD<sub>50</sub> (mice, iv; mg/kg): MT; 220, 6a; 110, 6b; 280, 7a; 60.

Table 1. (Continued)

Test organisms	9a	9b	10a	10b	11a	11b	12a	12b
<i>Staphylococcus aureus</i> 193	0.39	0.2	<0.2	<0.2	<0.2	0.39	1.56	0.78
<i>S. aureus</i> EMf	>100	50	>100	100	>100	100	>100	>100
<i>S. aureus</i> 209P	0.78	<0.2	0.39	<0.2	<0.2	<0.2	1.56	0.78
<i>S. aureus</i> MS 9610	>100	>100	>100	>100	>100	>100	>100	>100
<i>S. aureus</i> MS 9351	>100	>100	>100	>100	>100	>100	>100	>100
<i>S. aureus</i> MS 9861	0.78	0.39	0.39	0.39	0.39	0.39	1.56	1.56
<i>S. aureus</i> MS 10225	1.56	0.39	0.78	0.39	0.39	0.39	3.12	3.12
<i>S. aureus</i> MS 10246	>100	>100	>100	>100	>100	>100	>100	>100
<i>S. aureus</i> Smith	0.78	0.39	0.39	<0.2	0.39	0.78	1.56	1.56
<i>Micrococcus luteus</i> PCI 1001	<0.2	<0.2	<0.2	<0.2	0.2	<0.2	<0.2	<0.2
<i>Bacillus subtilis</i> NRRL B-558	1.56	0.39	0.78	0.39	0.39	0.39	3.12	3.12
<i>Corynebacterium bovis</i> 1810	0.78	<0.2	0.39	<0.2	0.78	<0.2	6.25	3.12
<i>Escherichia coli</i> NIHJ	25	3.12	1.56	3.12	1.56	1.56	6.25	6.25
<i>E. coli</i> K-12	50	6.25	12.5	6.25	12.5	6.25	50	25
<i>E. coli</i> K-12 R-5	100	12.5	12.5	12.5	12.5	6.25	50	25
<i>E. coli</i> K-12 ML 1629	>100	50	100	50	100	50	>100	100
<i>E. coli</i> K-12 ML 1410	>100	50	100	100	100	50	>100	100
<i>E. coli</i> K-12 ML 1410 R81	>100	50	100	100	100	50	>100	100
<i>E. coli</i> K-12 LA290 R55	100	12.5	25	12.5	25	12.5	100	50
<i>Klebsiella pneumoniae</i> PCI 602	3.12	3.12	3.12	3.12	1.56	3.12	6.25	3.12
<i>Shigella dysenteriae</i> JS 11910	1.56	0.78	0.78	0.78	0.78	0.78	1.56	0.78
<i>Salmonella enteritidis</i> 1891	6.25	3.12	1.56	3.12	3.12	3.12	3.12	3.12
<i>S. typhi</i> T-63	>100	50	50	50	50	25	100	50
<i>Enterobacter aerogenes</i> ATCC 13048	>100	50	50	50	100	50	>100	100
<i>Providencia</i> sp. Pv 16	>100	100	>100	>100	>100	>100	>100	>100
<i>Serratia marcescens</i>	50	12.5	25	12.5	50	12.5	25	12.5
<i>Proteus vulgaris</i> OX-19	100	25	12.5	25	50	12.5	100	50
<i>Pseudomonas aeruginosa</i> A3	50	3.12	6.25	6.25	25	12.5	12.5	6.25
Geometrical mean	18.58	5.68	7.63	5.97	8.43	5.96	20.01	13.47

Other 23-C-substituted analogs (**7a,b**~**12a,b**) were prepared similarly by treatment of **4** with ethyl-, butyl-, phenyl-, allyl-, vinyl-, and ethynyl-magnesium bromide, followed by deblocking. The faster-moving products on column chromatography (they are shown by attachment of "a" to the numbers) had, in all compounds including **6a**, small  $J_{14,23}$  values (2~3 Hz) in their  $^1\text{H}$  NMR spectra, and the slower-moving ones (shown by "b" after the numbers) including **6b**, larger  $J_{14,23}$  values (5.5~6.8 Hz). The faster- and the slower-moving products were, therefore, concluded to have the same spatial arrangement with those of **6a** and **6b**, respectively. **7a**:  $[\alpha]_D^{25}$   $-6^\circ$ , **7b**:  $+9^\circ$ , **8a**:  $+7^\circ$ , **8b**:  $+30^\circ$ , **9a**:  $+81^\circ$ , **9b**:  $+30^\circ$ , **10a**:  $+40^\circ$ , **10b**:  $-10^\circ$ , **11a**:  $-3^\circ$ , **11b**:  $0^\circ$ , **12a**:  $+65^\circ$ , **12b**:  $-39^\circ$ , all measured at  $c$  1 in  $\text{CHCl}_3$ . The structures of these compounds were confirmed by the elemental analysis.

Antibacterial spectra (Table 1) of these products were diversified, but the slower-moving products (minor products) always had, more or less, stronger activities than those of the corresponding faster-moving ones (major products). Among them, **8b**, **9b**, **10b** and **11b** were the most prominent. Acute toxicities (Table 1) of **6a**, **6b**, **7a**, and MT in mice suggest that the isomers having the same spatial arrangement with that of **6b** may have lower toxicity in comparison with the other series of isomers, respectively. Structure-toxicity relationships will be studied more in detail in the future.

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TSUTOMU TSUCHIYA  
SHUICHI SAKAMOTO  
NORIO KAJIKAWA  
SUMIO UMEZAWA

Institute of Bioorganic Chemistry,  
1614 Ida, Nakahara-ku,  
Kawasaki 211, Japan

MASA HAMADA  
HAMA O UMEZAWA

Institute of Microbial Chemistry,  
Kamiosaki, Shinagawa-ku,  
Tokyo 140, Japan

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